OSTEOARTHRITIS AS AN ORGAN DISEASE: FROM THE CRADLE TO THE GRAVE

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Abstract

Considered for decades as a cartilage disease, recent studies of osteoarthritis (OA) take us back to the concepts discussed at the naming of the disorder as “bone-joint-inflammation”. By describing the joint as an organ, can OA be called an organ disease – similar to heart disease? Is there a systemic (which system?) involvement? Would this help with diagnosis or therapy? Hyperplasia of the joint tissues is one of the most notable early features of the disease: articular cartilage thickens, chondrocytes proliferate and increase matrix biosynthesis, but not its incorporation; the subchondral bone densifies but is hypomineralised and there is an increase in bone marrow fat content. Associations between OA and hypertension, hypercholesterolaemia and blood glucose suggest systemic and metabolic components are involved. The source of pain is still unknown but there is evidence for peripheral and central sensitisation, joint deformity is difficult to quantify, but statistical shape modelling provides a tool to use as an imaging biomarker. A genome-wide association study meta-analysis has identified novel genes associated with hip shape with many genes related to tissue growth and development. There are associations between hip shapes and age of first walking as well as with obesity through adulthood. These life-course events and a recapitulation in old age of developmental processes suggest that the cradle may affect our path to the grave. These observations suggest that tissue regeneration approaches, treating only the cartilage in OA joints, may only be of limited benefit.

Keywords: Osteoarthritis, joint, systemic disease, metabolic disease, aetiology, growth, hyperplasia, development.

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of papers focusing on cartilage has not decreased, there is an increased number of papers that deal with other aspects of osteoarthritis pathology, including soft tissue interactions with cartilage and mineralised tissue.

Some forms of OA follow trauma and affect a single joint, an abnormal load on a normal joint. Most, however, are still classed as idiopathic – unknown cause – and are polyarticular in nature. Traditionally, idiopathic OA has been suggested to arise from normal loads on abnormal cartilage or from abnormal loads on normal cartilage. In both of these scenarios, the abnormality is often much more elusive than an obvious traumatic incident. Curiously, hand OA is one of the most prevalent forms of OA, affecting around 35% of adults (Haugen et al., 2011) and hands are rarely subject to abnormal loads, suggesting some other cause. Despite the history of the name described above, the emphasis in recent decades has largely been on the articular cartilage of the synovial joints as the affected tissue and biomechanical factors as the causative agents. Changes in other tissues have been treated as secondary, including alterations to subchondral bone and other tissues believed to arise from secondary inflammation and enforced inactivity.

In some recent publications, it is proposed that the joint should be considered as an organ (Loeser et al., 2012; Lories and Luyten, 2010; Radin et al., 1997). Can OA, therefore, be described as an organ disease in a similar way to heart disease? Is there a systemic involvement and, if so, by which system or systems? Does this help with diagnosis or therapy? Indeed, does this view complicate the treatment of OA? In this review the effects of OA on a number of joint tissues were examined and an attempt made to show that, although “cartilage first” cannot be ruled out, it is not the only, or even necessarily the best – explanation for the hyperplasia observed in many tissues forming the joint. Similarly, although biomechanics almost certainly plays an important role in the progression of the disease, its part in the incidence of primary generalised OA is less clear. It may be the factor that determines, in any individual, which joint displays the first signs; however, the working hypothesis was that generalised OA is a recapitulation or renewal of growth processes that should have ceased permanently at the end of development and that these are driven by metabolic and systemic factors.

It is proposed that OA, at least in its generalised form, is a systemic musculoskeletal disease, further developing ideas presented previously (Aspden, 2008; Aspden, 2011; Aspden et al., 2001). Specifically, it is suggested that OA is characterised by dysregulated growth of musculoskeletal tissues and that many of the cells appear to have reverted to an earlier developmental phenotype. The outcome is new tissue being formed in the wrong place and at the wrong time, matrix components not being properly incorporated and a resulting loss of the mechanical properties required for normal function. Initially a discussion of articular cartilage is presented, as this is largely the focus of tissue repair strategies, before considering some of the other tissues comprising the joint.

**Articular cartilage**

Cartilage loss is one of the most obvious radiographic signs, often described as “joint space narrowing”. On a radiograph the bone is opaque and the cartilage is transparent. If this remarkably smooth load-bearing surface is lost, joint function will be compromised as the bones are forced closer together and ultimately touch – rubbing together. Several significant structural changes result, such as the loss of the low friction environment, increased sensations of pain as innervated bones rub together and, therefore, changes in the bone and functional impairment. The loss of cartilage, however, is a late event that occurs after more subtle structural and molecular changes in both the bone and the articular surfaces and is accompanied by a chronic, low-grade inflammation. This is in direct contrast to the pathophysiology of RA where the primary initiating factor is inflammatory in nature and the changes in cartilage and bone are secondary (Thysen et al., 2015).

The role of the chondrocyte in articular cartilage is extensively reviewed (Goldring, 2000b; Goldring and Goldring, 2007). Essential features of the process often described as degeneration are tissue swelling, cell proliferation, fibrillation and finally erosion. The first observable sign is swelling of the tissue as it absorbs more water (Maroudas, 1976; Wang et al., 2008). This is accompanied by renewed cell division (Mankin et al., 1981), or cloning, of the chondrocytes within their chondron, producing multiple cells where in normal tissue there are typically only 2-4 chondrocytes (Mankin et al., 1981; Poole, 1997). Synthesis of the principal matrix molecules increases (Goldring and Goldring, 2007), including collagen type II (Aigner et al., 1997) and aggrecan (Roughley and Mort, 2014). In the very early stages of the disease, when only minor surface fibrillation is apparent, there is substantial release of aggrecan fragments. However, the net tissue concentration of aggrecan does not decrease (reviewed by Heinegard and Saxne, 2011), indicating that much of this new material appears to be lost rapidly to the joint cavity. Hence, one of the markers for increased tissue turnover is elevated levels of aggrecan fragments in the synovial fluid (Lohmander et al., 1999; Roughley and Mort, 2014). There are also reports of substantial increases in the abundance of cartilage oligomeric protein (COMP) and cartilage intermediate layer protein CILP-1. These new accumulations occur close to the chondrocytes (Lorenzo et al., 2004). It is reported that the altered distributions of these two proteins present a typical and distinct hallmark of the cartilage in the early osteoarthritic
process. This is the result of molecules being lost from one compartment, through degradation, and accumulating in another as a result of new synthesis (Heinegard and Saxne, 2011). It is also reported that this distinctive distribution is similar to that found during embryonic development (Heinegard and Saxne, 2011; Shen et al., 1995). Interestingly, the expression of collagen type II changes from type IIB, the adult form, to type IIA – a splice-variant that is normally expressed during development (Aigner et al., 1999). Work from Bruce Caterson’s lab shows that there are specific epitopes of chondroitin sulphate (4-C-3, 7-D-4 and 3-B-3(-)), more commonly found in foetal tissue, decorating the cell surface proteoglycans on activated stem cells. This could be a signal that these cells are a target for tissue development (Hayes et al., 2018). The significance of these specific epitopes is that they are commonly found on progenitor cells, which occur in increased numbers in OA tissue. This provides further evidence of a reversion to a more developmental-like phenotype. Increased matrix synthesis is accompanied by increased biosynthesis of matrix metalloproteases (MMPs), prostaglandins and other inflammatory factors. These may arise from elevated levels of interleukin IL-1 and tumour necrosis factor TNFa (Goldring, 2000a; Goldring, 2000b; Zhuo et al., 2012). An increase in the pro-inflammatory precursor arachidonic acid, and other lipids, is also reported to be associated with histological severity of OA (Lippiello et al., 1991). Finally, the tidemark – which marks the junction between calcified and uncalled cartilage – is duplicated and advances into the previously uncalled tissue (Oettmeier et al., 1989; Radin et al., 1991). This is another indicator of renewed growth and tissue formation, though the underlying mechanisms are still unknown. A stereological study reports an increase in the area covered by the tidemark, with a greater number of blood vessels crossing it, that is taken to be indicative of a vascular involvement in the pathology (Bonde et al., 2005). This feature is quite difficult to model in animals due to the differences in OA pathology of cartilage structure, particularly in rodents. However, there is an in vitro co-culture model of osteoblasts and chondrocytes that can be used to model the changes in the tidemark (reviewed by Thysen et al., 2015) and a model in dogs (Daubs et al., 2006).

Articular cartilage may be considered as a biological fibre-composite material in which collagen fibrils provide tensile reinforcing to a highly hydrated proteoglycan gel (Aspden, 1994). Viewed in this way, it is the interactions between collagen and the other components that provide mechanical integrity (Aspden, 1994; Burgin et al., 2014; Hukins et al., 1984; Lewis et al., 1998) – not simply a stiff collagen ‘network’ and a filler. The organisation of the fibrils (Aspden and Hukins, 1981; Jeffrey and Aspden, 2006), the electrostatic properties of the gel (Dean et al., 2003; Seog et al., 2002) and interactions between the fibrils and the gel (Aspden, 1994; Rojas et al., 2014) are tightly regulated during development and in response to a changing loading environment to produce appropriate mechanical properties. Proteoglycan turnover, but not collagen turnover (Heinemeyer et al., 2016), then maintains the mechanical stability of the tissue. The carbon-14 bomb dating method used by Heinemeier et al. (2016) demonstrates the relative stability of the collagen structure in both healthy and diseased tissue. Page Thomas and colleagues demonstrate, in rabbits, that – after an immediate extensive loss of proteoglycans from cartilage – there is suppression of the production in the immediate aftermath of injury but this then doubles its rate after one week (Page Thomas et al., 1991). Excess synthesis of new matrix and a reversion of the chondrocytes to a developmental-like phenotype could weaken the tissue by inappropriate remodelling. Weakening of the tissue in this way could then make it susceptible to mechanical damage. Even in elderly tissue, chondrocytes respond to mechanical (Plumb and Aspden, 2005) and chemokine stimuli (Plumb et al., 2006) and to tissue damage (Jeffrey et al., 1997). However, once the tissue structure, especially the collagen organisation, has been lost it does not presently seem possible to restore it. Catterall and colleagues show that the half-life of collagen is very long, in excess of 100 years; therefore, the loss of collagen appears to be irreversible (Catterall et al., 2016). Studies of mechanical loading, on elderly human articular cartilage, indicates that both cyclic and static loads inhibit matrix macromolecule biosynthesis (Plumb and Aspden, 2005) and could over-ride the normally stimulatory effect of IGF-1 (Plumb et al., 2006; Wheeler et al., 2009). This differs from results from young bovine tissue where cyclic load is generally found to be stimulatory (Guilak et al., 1994; Larsson et al., 1991; Palmoski and Brandt, 1984; Sah et al., 1989). Supporting this, preliminary gene expression studies comparing OA cartilage with that obtained from hip fracture patients indicated that cyclic loading produced no changes in gene expression for matrix macromolecules at greater than a 4-fold level but that there were changes in the expression of growth factors (FGF-18, FGF-2, VEGF and WNT16), all of which promote chondrocyte division, and molecules associated with increased matrix turnover such as COX-2 and ADAMTS-1 (Meeting abstract: Plumb and Aspden. Osteoarthritis Cartilage 2005 13: S104.).

**Bone**

A number of signs of OA appear in the bones forming the joint. These include sclerosis of the subchondral bone, the formation of so-called “cysts” in the trabecular bone, and osteophytosis. Cysts are cavities in the cancellous bone, often found in late-stage OA below the loaded articulating region (Havdrup et al., 1976). The mechanism of cyst formation is still unclear. Recent studies favour the idea of bone contusion over synovial fluid intrusion, due to their
association with bone marrow lesions seen using magnetic resonance imaging (MRI) (Carrino et al., 2006; Crema et al., 2010). Irrespective of how they are initiated, removal of so much bone suggests significantly increased osteoclastic activity (Havdrup et al., 1976). In contrast, osteophytes are outgrowths of bone and cartilage found in many patients at the margins of diarthrodial joints or as outgrowths in the central portions of the articular space in about 15% of patients (McCauley et al., 2001), as well as in the spine, around zygoapophyseal joints and vertebral bodies particularly in the lumbar region (Klaassen et al., 2011). Osteophytes form by endochondral ossification (Gelse et al., 2003). The latter process shows many similarities to bone formation during development and the signalling pathways activated during osteophyte formation are shown to be similar to those found in callus formation during fracture repair (Arden and Nevitt, 2006; Patel et al., 2003; van der Kraan and van den Berg, 2007). Bone usually adapts in response to mechanical loading and growth of new tissue, in the absence of such a stimulus, requires other causes. One suggestion is that TGF-β could provide this signal (Scharstuhl et al., 2002; van Beuningen et al., 1994). Similarly, subchondral sclerosis has traditionally been regarded as a reaction to increased load following cartilage loss. Several studies, however, indicate that bone changes are evident concurrently with cartilage fibrillation (Dedrick et al., 1993) or that sclerosis is evident radiographically before joint space narrowing (Buckland-Wright et al., 1995).

Patients with radiographic OA of the hip are reported to have an elevated bone mineral density not only in the hip but also in the distal radius, vertebrae and calcaneus (Nevitt et al., 1995). Scintigraphy, using technetium 99m, shows increased bone forming activity in joints with pathological signs of OA (Buckland-Wright et al., 1995). Laboratory studies show alterations in the bone matrix and in osteoblast behaviour. In the hip, an increase in cancellous bone volume of about 60% was found but this was associated with a reduced mineralisation (Li and Aspden, 1997a). Using back-scattered as well as secondary emission electron microscopy, the appearance of the cancellous bone was found to be similar to woven bone seen in fracture repair (Li et al., 1999) with evidence of osteoclastic resorption in the form of Howship lacunae. In addition, although the subchondral bone plate was thicker, it too was less well mineralised (Li and Aspden, 1997b) (Fig. 1). Increased amounts of bone are reported in the iliac crest of patients with OA of the hand (Gevers et al., 1989a; Gevers et al., 1989b), together with greater levels of growth factors (Dequeker et al., 1997) – supporting the concept that OA may be a part of a generalised disorder. Among the anabolic factors identified as important in bone formation

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**Fig. 1.** Scanning electron micrograph of subchondral bone from a 75-year-old patient following total hip replacement for OA. The porous and coarse texture of the bone matrix is apparent, and measurement showed the matrix to be hypomineralised.
are the Wnt family and several studies show their differential regulation in OA (Burr and Utreja, 2018; Corr, 2008; Luyten et al., 2009). In a preliminary gene expression study, not only was differential regulation of some of the more obvious members of Wnt signalling pathways found but that the most differentially regulated gene was that for NHERF1 (Sodium/hydrogen exchange regulatory cofactor 1; SLC9A3R1) that was strongly down-regulated (Hochet and Aspden, 2007). NHERF-1 acts with NHE-3 (sodium/hydrogen exchanger 3, SLC9A3) to regulate intracellular pH through control of sodium and potassium ion concentrations but is also shown to interact with PTEN (phosphatase and tensin homologue), a tumour suppressor, and to regulate cell proliferation in various cancers and cell types (Li et al., 2015; Molina et al., 2012; Takahashi et al., 2006). Immunofluorochemistry studies demonstrate cytoplasmic accumulation of NHERF-1, PTEN and β-catenin, consistent with increased AKT (also known as protein kinase B) activation and Wnt signalling due to the translocation of the β-catenin that has accumulated in the cytoplasm to the nucleus in both chondrocytes and osteoblasts from OA femoral heads (Griffin-Walker et al., 2017; White et al., 2015). A similar translocation was found from cell membrane to the cytoplasm for NHERF-1 to that which is reported in tumour growth, supporting its possible role in hyperplasia (Griffin-Walker et al., 2017).

**Obesity and adipose tissue**

There is a recognised association between being overweight and OA of knees, hips and hands (Issa and Griffin, 2012; Reyes et al., 2016; Thijssen et al., 2014). Is this mechanical or metabolic, or both? Being overweight is known to predate the incidence of disease (Felson et al., 2000). Results from both the Chingford and the Framingham studies show that body mass index (BMI) is linked to all patterns of knee OA (Cooper et al., 1996; Felson et al., 1988; Hart et al., 1995). Mechanical overloading is the obvious mechanism for subsequent cartilage degeneration. This may play a role in knee and hip OA but much harder to understand in OA of the hand, where baseline obesity is associated with the incidence of hand OA 23 years later (Carman et al., 1994). A growing literature is increasingly pointing away from a simple link between mechanical overloading and OA. Or, conversely, that obesity is a consequence of reduced activity due to joint pain. The role of adipose tissue as an endocrine organ secreting adipokines is increasingly prevalent (Aspden, 2011; Berenbaum and Sellam, 2008; Francisco et al., 2018; Issa and Griffin, 2012; Pottie et al., 2006; Reyes et al., 2016; Thijssen et al., 2014). The association between OA and being overweight is strongest with incidence and severity but not with progression. This is shown clearly in a study that recruited morbibly obese women (body mass index (BMI) between 30 and 50 kg/m²), on the basis that they would progress fastest and enable a therapeutic approach to be tested most rapidly (Hollie Le Graverand et al., 2009). Joint space narrowing does not increase with BMI and they conclude that there is nothing to be gained from using this measure by recruiting obese individuals.

Following observations that femoral heads recovered from OA patients, after total hip replacements, seem to contain much more fat than those from osteoporotic patients, the fat content and the fatty acid composition was measured (Plumb and Aspden, 2004). It was found that the bone cores from OA bone contained twice the mass of fat per unit volume of tissue as osteoporotic bone. The fat contained more (n-6) fatty acids, and the amount of arachidonic acid was double that found in the tissue from osteoporotic bone. These fatty acids are the substrates for the cyclo-oxygenase enzymes, that are the target of many non-steroidal anti-inflammatory drugs, and result in the formation of prostaglandins – active pro-inflammatory mediators. Features of OA are suggested to have many similarities with atheromatous vascular disease (Marks and Allegrante, 2002) and the role of lipids in calcification is an active area of study in the formation of atherosclerotic plaques (Parhami et al., 2001), as well as the differentiation of stem cells into osteoblasts and adipocytes (Parhami et al., 1999). These systemic and metabolic markers indicate that OA may, therefore, be a part of the so-called Metabolic Syndrome (MetS). MetS has a very broad spectrum of included syndromes/diseases and includes obesity, hypercholesterolaemia, hyperlipidaemia, hypertension and cardiovascular disease as characterised by atherosclerotic plaques (Zhuo et al., 2012). However, a recent systematic review of the link between MetS and OA in different joints, by Li and Felson, indicates that there is no relationship with hip OA. The possible associations with hand and knee OA are not clear, due to limitations in the studies. These are reported to be small in participant numbers, mainly cross-sectional in nature with few outcome measures or adjustment for co-variates (Li and Felson, 2018). There is a growing understanding that cells of mesenchymal origin can not only be directed to differentiate down specific lineages, by soluble factors activating the relevant nuclear receptor, but also may be redirected by modulating the nature of their stem-cell niche. Hence, modulating the stem-cell precursors may affect all the cell lineages derived from those stem cells and, consequently, affect the composition and, therefore, the functionality of all the derived tissues. This knowledge begins to open new avenues for understanding why so many tissues are affected by the disease process and, perhaps, why adipose tissue is one of the key drivers of OA (Aspden, 2011). Genetic and epigenetic factors are shown to be transmitted through cell lineages. For example, re-expression of growth and development factor 5 (Gdf-5), is found in the synovium of mouse models
of OA, with a role in cartilage repair. Also, a second gene, one of the bone morphogenetic factors Bmp-7, is shown to play a critical role in joint patterning (Roelofs et al., 2017).

**Synovial hyperplasia**

One aspect of osteoarthritis development which has received a significant amount of interest is synovial hyperplasia. A review by de Lange-Brokaar and colleagues uncovers 100 different papers that show different aspects of synovial hyperplasia in humans (de Lange-Brokaar et al., 2012). This includes investigations of synovial tissue proliferation, infiltration of the tissue by macrophages and – to a lesser extent – neutrophils, quantification of cytokines such as IL-1β and IL-6, the role of other immune cells such as T-cells and the degree of inflammation that is involved. A PubMed search using the terms “synovial hyperplasia” and “osteoarthritis” showed that there were over 200 papers on this particular topic, indicating the complexity of the synovial tissue and its role in OA pathophysiology. It is commonly believed that there is no inflammatory involvement in the development of OA. However, this view seems to have shifted to show that there are indeed low levels of inflammation present in many patients with OA, albeit at a lower level than is commonly associated with RA (Sokolove and Lepus, 2013). There are reports of increased levels of the cytokines IL-1β and TNF-α (tumour necrosis factor alpha) in cartilage matrices undergoing degeneration (Kapoor et al., 2010). Many papers highlight that in OA tissue there are increased features of hyperplasia, demonstrated by increased scores compared with normal tissue. The seminal work of Mankin, in developing a robust scoring system for the classification of hyperplasia and associated changes observed in OA (Mankin et al., 1981), cannot be overlooked and has been key in changing the mindset that the condition of the cartilage is the most critical pathophysiological change in OA development. This is reflected in the subsequent updates to the consensus OARSI scoring system (Pritzker et al., 2006).

**Other indicators of growth**

The joint capsule is a tough, strong tissue that not only contains the joint fluid but also plays a vital, and commonly under-estimated, role in the biomechanics of synovial joints. Capsular ligaments augment other ligaments to maintain joint stability (Ralphs and Benjamin, 1994). In OA there is progressive fibrosis of the capsule and of the ligaments, resulting in thickening and shortening of the tissue and a reduction in its flexibility (Lloyd-Roberts, 1953). Ruptures of the anterior cruciate ligament are found to be far more common in subjects with symptomatic knee OA (Hill et al., 2005). In patients, a history of knee injury is associated with increased radiographic OA (Johnson and Hunter, 2014). Although such an injury is a known risk factor for secondary OA, it cannot be determined which came first. However, there is evidence from the Osteoarthritis Initiative that loss of ACL integrity and the development of incident radiographic OA are not related (Johnson et al., 2015). It is possible, in some patients, that ligament damage arose after the onset of disease as a consequence of altered tissue biomechanical properties. Studies of OA in the STR/ort mouse show that cruciate ligament metabolism is upregulated before radiological signs are present and that the tissue is weaker than in controls (Anderson-MacKenzie et al., 1999; Staines et al., 2017).

The changes in quantity, appearance and viscosity of synovial fluid in OA are well documented but high numbers of mesenchymal precursor cells (mesenchymal stem cells (MSCs)) are reported in synovial fluid from OA patients (Jones et al., 2004). Because levels are higher than in RA, it is suggested that these are not simply a consequence of an inflammatory process. These cells are pluripotent, with their ultimate phenotype able to be manipulated by altering the chemical composition of the culture medium. They possess the ability to be differentiated in culture to three main cell types: osteoblasts, chondrocytes and adipocytes. The origin of the MSCs is currently unknown, but cells with stem-cell-like properties are being identified in most joint tissues (Franceschetti and De Bari, 2017). Multiplication of stem cells could have a key role in the underlying tissue hyperplasia being described here as well as in the return to a more developmental phenotype but their presence, and in what numbers, is still an active area of investigation. By better understanding the origin of these cells and the factors required to drive them to differentiate into specific cell lineages, a deeper explanation of why cells in OA tissue seem to revert to a more developmental phenotype may be gained.

Among many possible growth, genetic and epigenetic factors that associate with OA incidence and progression, two warrant special mention; so-called axon guidance molecules – because they are normally associated with development – and cartilage oligomeric matrix protein (COMP) – because it is widely used as a serum biomarker. In preliminary studies, using gene arrays, an approximately 2-fold higher expression of spondin-1 (F-spondin) was found in OA bone than in osteoporotic bone and further exploration revealed increased expression of neuropilin and semaphorins (unpublished). Another study reports a 7-fold increase in spondin-1 in human OA cartilage (Attur et al., 2009). These molecules are traditionally studied during embryonic development and are associated with growth of nerves into newly formed tissues. Studies in embryonic cartilage suggest that spondin-1 has the capacity to enhance chondrocyte terminal differentiation and mineralisation through interactions in its thrombospondin-repeat domain.
and TGF-beta dependent pathways (Palmer et al., 2010). These studies provide supporting evidence for a recapitulation of developmental processes occurring in diseased tissues.

Finding biomarkers for OA is a high priority, as evaluating radiographic joint space – which requires imaging – is the only current biomarker accepted by the US Food and Drug Administration (FDA) and is not a very sensitive method. COMP is identified as a cartilage matrix protein (DiCesare et al., 1995) and it was soon recognised that its degradation in OA could reflect disease processes occurring in the matrix (DiCesare et al., 1996). Elevated levels of COMP are found in the serum of patients with OA and it is proposed as a biomarker for diagnosis and prognosis (Verma and Dalal, 2013). Much is reported of attempts to elucidate its roles within cartilage but, for the purposes of this review, of considerable interest is the finding that COMP is expressed in the apical ectodermal ridge (AER) of the human embryo at gestational week 8, as well as in the cavitating joint at weeks 10 and 12 (Koelling et al., 2006). The AER is the starting point of limb formation and, hence, it appears to be a key player in the formation of the limbs. This is reinforced by another study in which COMP is found to be involved in the regulation of endochondral bone growth (Kong et al., 2010). Perhaps this makes it less surprising that elevated levels are found in OA joints, if there is a reversion to an earlier developmental phenotype.

**Infrapatellar fat pad**

The infrapatellar fat pad, located distally to the patella and the femur is shown to be a localised source of adipokines in OA as well as functioning as a buffering and lubricating tissue (Favero et al., 2017). Clockaerts et al. argue, in a narrative review of the area, that the infrapatellar fat pad should be considered to be an active OA tissue. This is due to the cytokines produced, the innervation which may be linked to OA pain, and the production of adipokines that inhibit the biosynthesis of cartilage matrix proteins (Clockaerts et al., 2010). In a subsequent paper, however, they show that conditioned media from the infrapatellar fat pad inhibited catabolic processes in the cartilage (Bastaanssen-Jenniskens et al., 2012). Han and colleagues go a little further and start to speculate as to whether the role of fat localised to the joint area is a good or bad thing (Han et al., 2014). They show, in MRI-based studies, that there were several significant changes in the infrapatellar fat pad that had negative consequences for the development of OA, mainly related to volume, joint space narrowing and any involvement of bone marrow lesions. Clearly, the significant changes which are reported in the infrapatellar fat pad add to the idea of the joint being an organ. Another paper shows that inflamed infrapatellar fat pads are shown to be hyperintense on MRI scans of the knee. When correlated with structural changes, this is found to be associated with worsening of bone marrow lesions, a feature of OA pathology and more generalised joint damage (Jarraya et al., 2017).

**Joint shape and OA**

In addition to molecular events, the understanding of how anatomical features, such as joint shape, influence disease pathophysiology has evolved from simple gross observations. Malformation of a joint during development, e.g. developmental dysplasia of the hip, is a known risk-factor for later OA. However, more recently, it has been shown that less overt changes – often found associated with demanding sporting activities in adolescents – appear, at least in part, to underlie femoro-acetabular impingement – now recognised as a cause of later OA (reviewed by Pun et al., 2015). But what about more subtle variations in joint shape? Can these be measured and do any of the features identified relate to OA incidence or progression? It has been shown that the radiographic outline of the femoral head of patients with hip OA has a different shape from that of matched asymptomatic controls (Gregory et al., 2007) (Fig. 2). This shape can be quantified using Statistical Shape Modelling (SSM) (Cootes et al., 1995), which uses principal component analysis to describe the shape in terms of independent “modes of variation”. Each image in a dataset then receives a score for each mode, which describes how many standard deviations it lies from the overall mean of all the images. Using radiographic images from the Rotterdam study, none of whom were symptomatic at baseline, it was found that a number of modes were significantly associated with OA severity six years later and those who progressed most rapidly to hip replacement could be identified as a sub-group (Gregory et al., 2007). Subsequently, using SSM from DXA (dual-energy x-ray absorptiometry) images, it was shown that measures of hip shape were associated with radiographic hip OA, and to a lesser extent hip pain, in the MrOS (osteoporotic fractures in men) cohort (Faber et al., 2017). The method showed promise as an imaging biomarker, as it
was found that statistically significant changes could be measured in shape over a 12-month period in a small longitudinal cohort comprising 62 individuals (Barr et al., 2018). Using data from the MRC National Survey of Health and Development (a birth cohort all born in one week in 1946), it was found that life-course factors were associated with the shape of the hip and spine at age 60-64 years as determined by DXA imaging. Being overweight, especially greater gains in weight during life, were associated with a flatter femoral head and a shorter femoral neck (Muthuri et al., 2017). Other epidemiological studies, using the same cohort, show that knee OA is associated with high BMI throughout adult life (Wills et al., 2012) and that hand OA is associated not only with high BMI but also with low birth weight, but only in men (Sayer et al., 2003). Intriguingly, it has recently been found that age at first walking is associated with hip shape (Ireland et al., 2018) as well as with bone strength at the hip (Ireland et al., 2017). Using images from the ALSPAC (Avon Longitudinal Study of Parents and Children) cohort, the association of genes already identified as related to OA with hip shape modes was explored and several susceptibility loci were found, including KLHDC5-PTHHLH (Kelch-like family member 5-parathyroid-like hormone), DOT1L (DOT-1 like methyltransferase) and COL11A1 (Collagen 11Alpha1) (Baird et al., 2018). Combining these data with those from other cohorts, using a meta-analysis, has identified 8 SNPs independently associated with hip shape and, curiously, many of them were strongly related to the process of endochondral bone formation (Baird et al., 2018b). Studies of SSM in the Tasmanian Older Adult cohort (TASOAC), with multiple imaging modalities, had several modes of variation determined by SSM that were associated not only with radiographic hip OA, hip cartilage volume and muscle strength but also with pain and effusion-synovitis (Ahedi et al., 2017). Further analysis of these data has shown that hip shape as defined by SSM is also a predictor for the risk of total hip replacement (Mezhov et al., 2018).

Conclusions and clinical implications

Evidence is mounting that OA is an organ disease affecting the whole joint and its component tissues (Fig. 3). Changes in muscle, nerves and pain sensitivity, ligaments and the vascular system have all been documented and were reviewed previously (Aspden, 2008). The working hypothesis is that generalised OA affects the whole musculoskeletal system because it arises from renewed, dysregulated, growth of tissues derived largely from cells from the mesenchymal lineage. The particular joint or joints affected may then be determined by subtle biomechanical factors such as joint shape. Evidence suggests a reversion of the phenotype of their constituent cells to an earlier developmental stage that results in:

- faster turnover and hyperplasia of bone,
- increased adiposity leading to obesity,
- fibrosis of ligaments and joint capsule,
- altered muscle phenotype possibly including fat deposition and muscle weakness,
- proliferation of cartilage chondrocytes,
- increased matrix synthesis, but not incorporation, all leading to weakened tissue and mechanical breakdown. Identifying alterations in mesenchymal stem cells and their niches may provide new insights into the proliferation of these tissues. The particular joints affected most may be dependent on local biomechanical factors and statistical shape modelling is beginning to suggest there may be subtle variations in hip shape that act throughout the life-course. However, the underlying disease, it is believed, is metabolic and systemic (Aspden et al., 2001). Some of these pathological changes have been recognised before and the Oxford surgeon Josep Trueta commented, “The osteoarthritic process thus appears to be an attempt to transform a decaying joint into a youthful one” (Harrison et al., 1953). After half a century of focussing on articular cartilage, to
the almost complete exclusion of other joint tissues, there is a returning again to the idea of OA as a joint disorder and a beginning to the understanding that processes throughout our life-course, from the cradle to the grave, may play a role in the pathogenesis of OA. A renewed focus on bone and the soft tissues of the joint in tandem with the cartilage will give a much more balanced picture of subtle changes that characterise OA.

These observations have many clinical implications. It becomes essential to identify OA in its early stages, especially those who are going to progress most rapidly (Gregory et al., 2007). Such methods and the resulting biomarkers are also essential for the development and effective testing of Disease Modifying OA drugs (DMOADs) in relatively short periods of time. Identifying circulating or urinary factors, or combinations of factors, deriving from tissues other than cartilage, such as bone, muscle or fat, may also provide new biomarkers of disease. Better still, a combination or panel of biomarkers, circulating, excretory and imaging, may harness better the markers we already know and provide a more sensitive measure of progression. It is believed that tissue engineering approaches to cartilage repair in osteoarthritis are almost certain not to succeed, at least in the short to medium term. If the natural tissue has been destroyed and the underlying problem has not been addressed, what chance is there for implanted tissue to survive, especially as, at least initially, the engineered tissue is commonly biomechanically inferior to the original? The exception would be local repair of damaged cartilage following trauma where it may be possible, with the correct combination of mechanical and biochemical cues, to engineer a tissue replacement that would be strong enough and could integrate with the existing tissue.

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Editor’s notes: There were no questions asked by reviewers for this paper therefore there is no Discussion with reviewers section. The Scientific Editor responsible for this paper was Martin Stoddart.